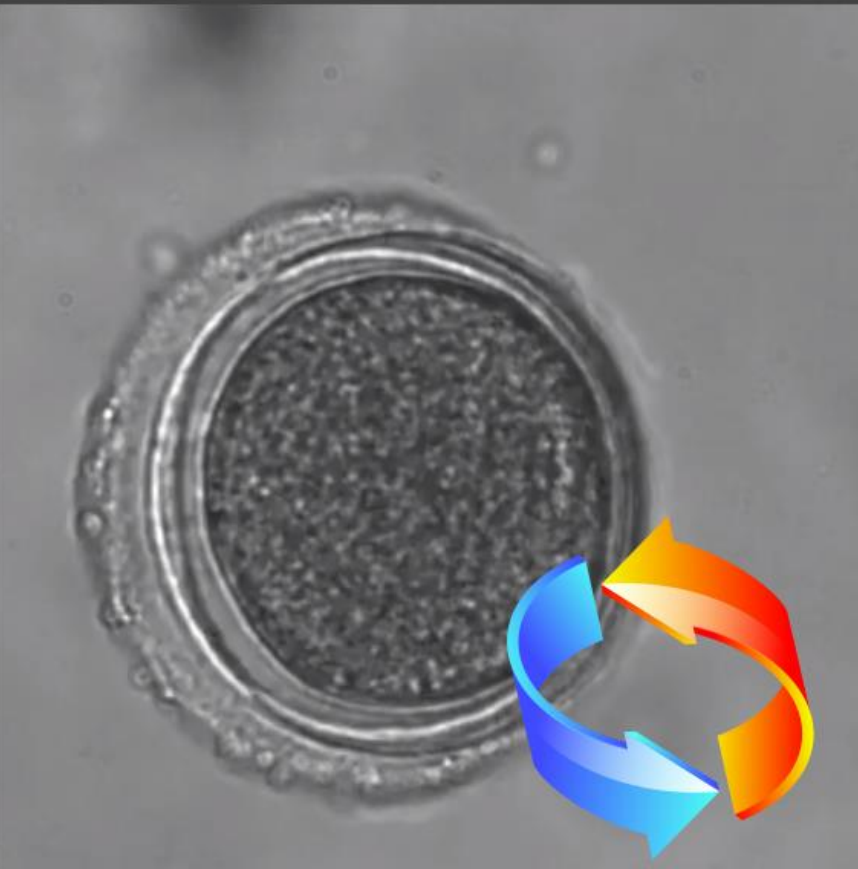


TRACKOPT - KICKOFF

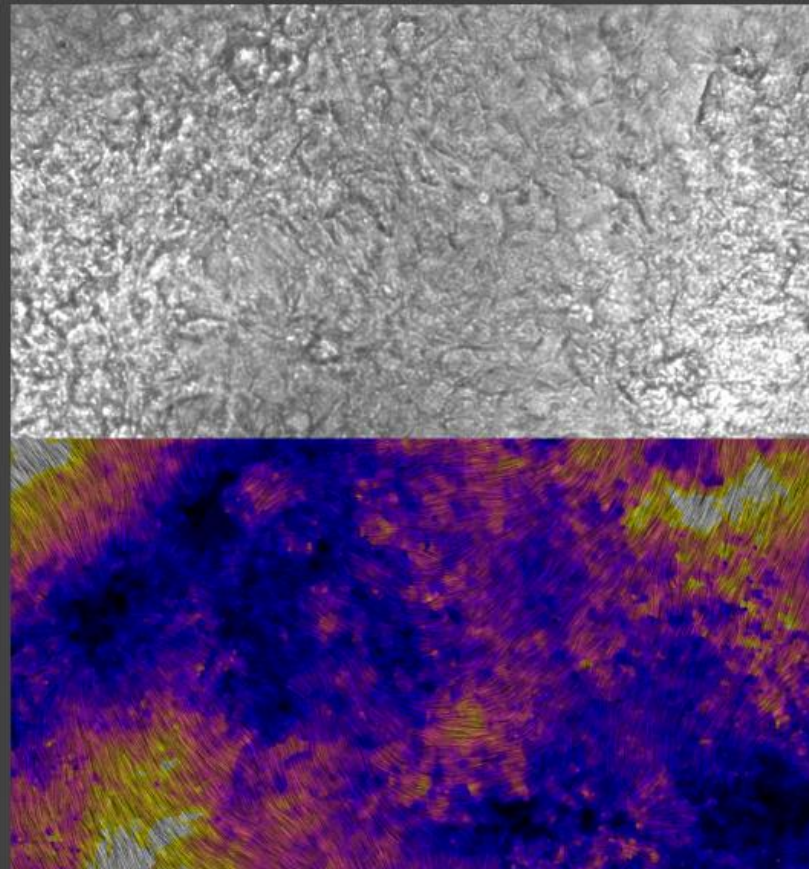
Ivo Ihrke
Universität Siegen

TRACKOPT – APPLICATION CELL TRACKING

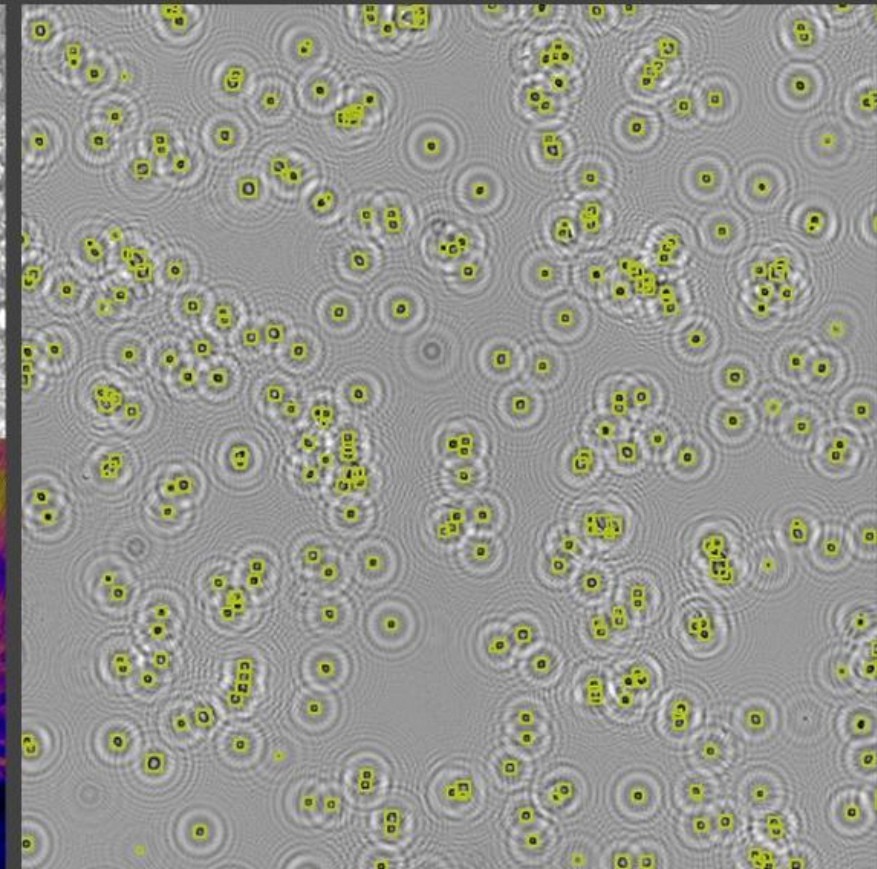
Fall I: Festkörpertransformation



Fall II: elastische Verformung



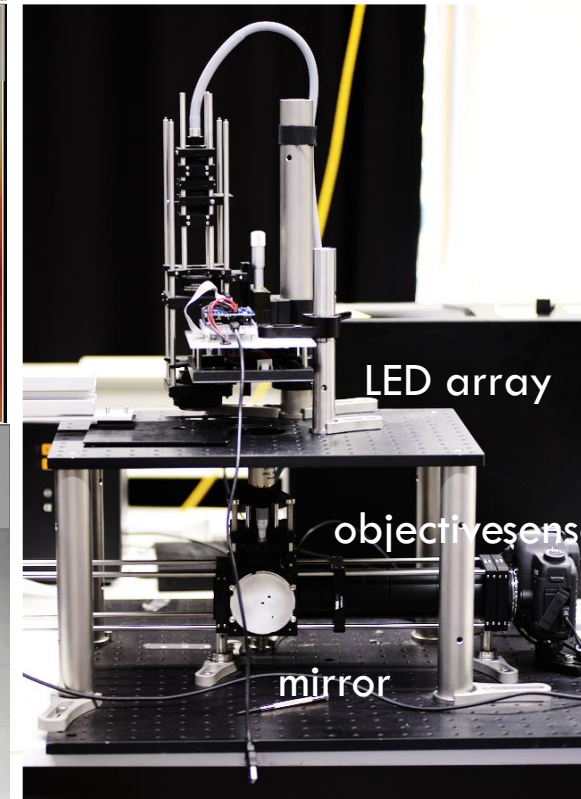
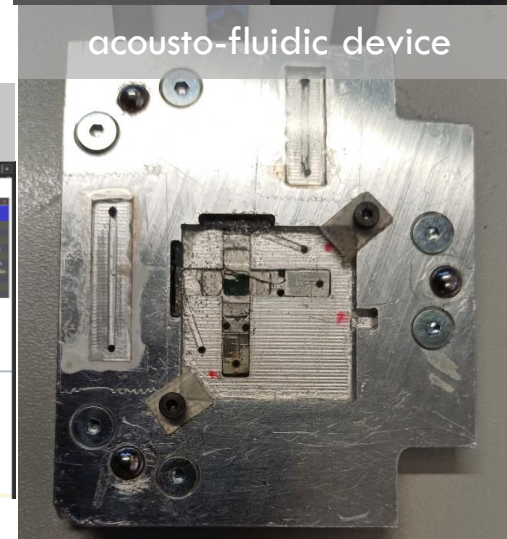
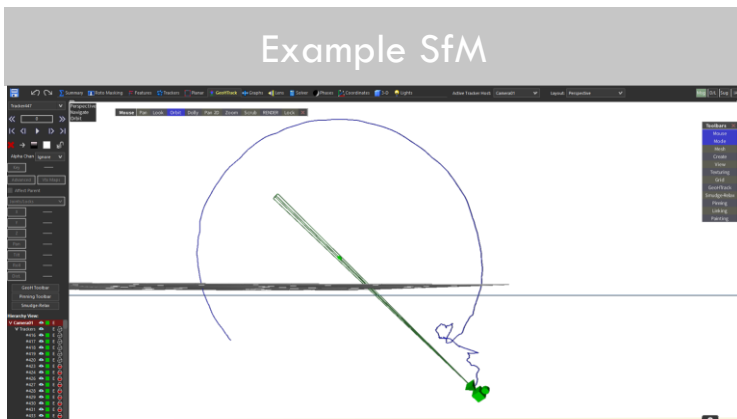
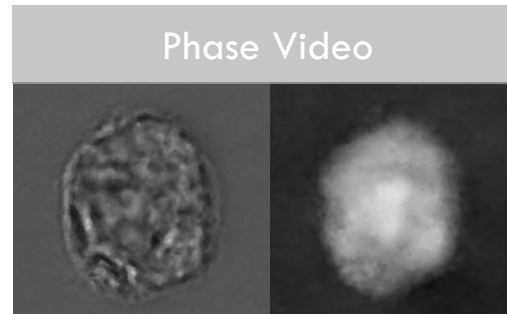
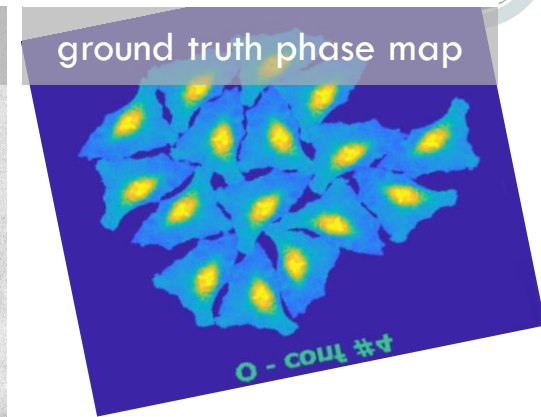
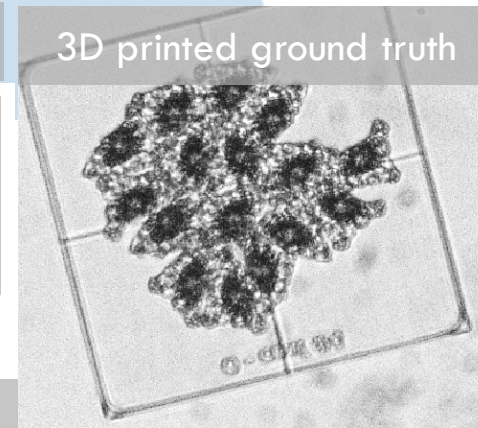
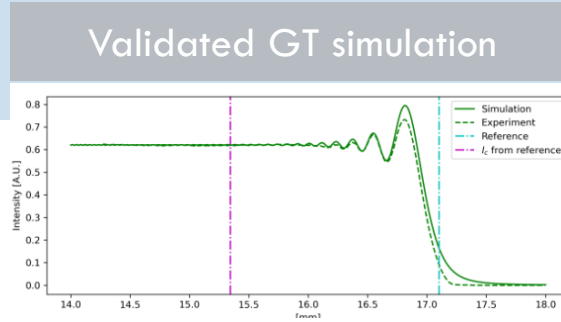
Fall III: holografische Partikel



CASE I: RIGID BODY MOTION

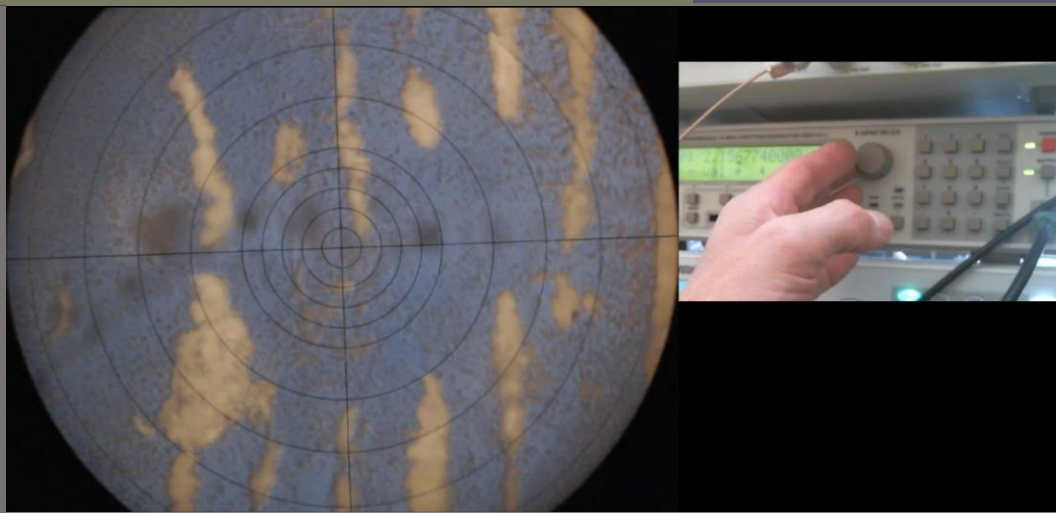
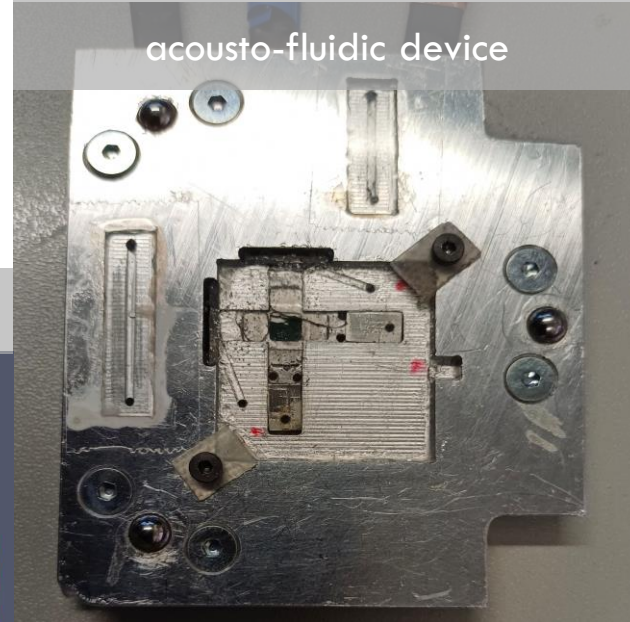
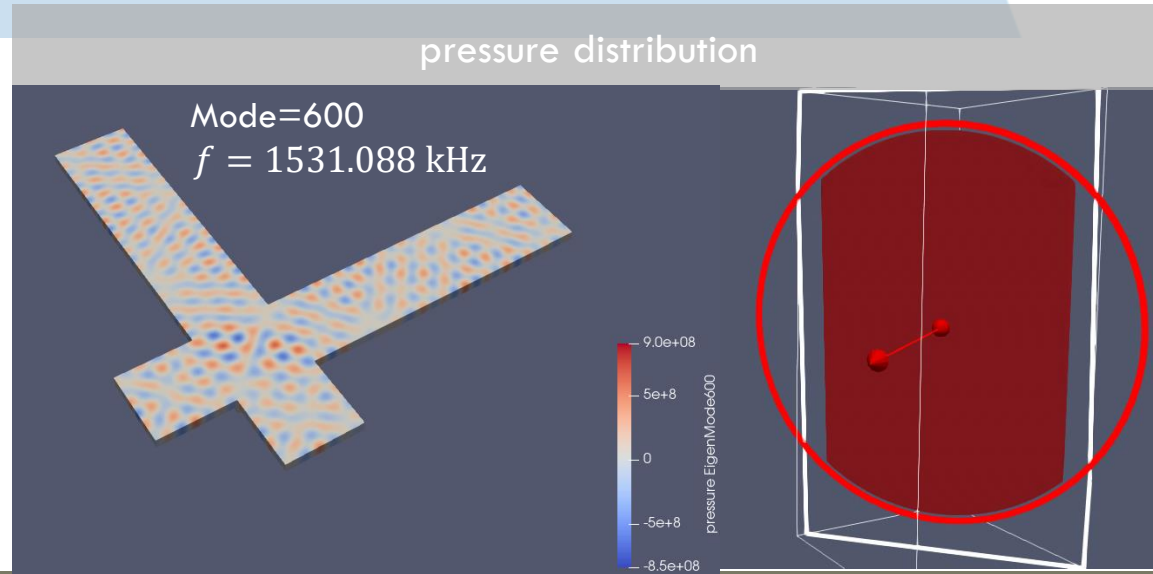
Data has been/will be acquired in a concurrent project (L2S 3D Microscopy)

- **Goal:** pose estimation of single cells / cell clusters
 - tracking to enable structure from-motion
- Challenge: image formation - PDE
- Properties:
 - Periodic motion
 - 3D constraints

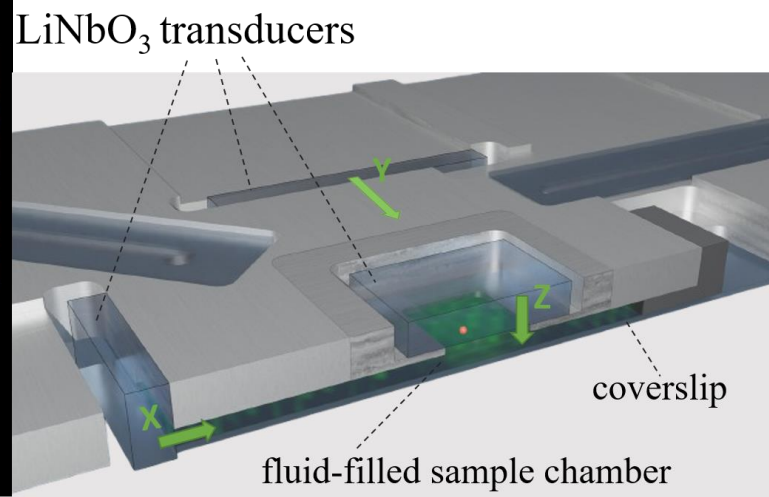


MICROFLUIDIC MANIPULATION DEVICE

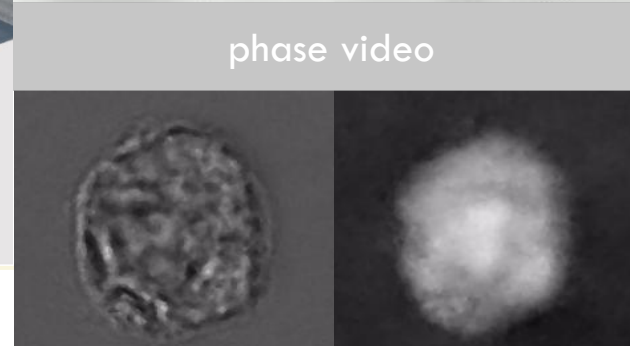
- Enables touchless manipulation (translation, 2 rotation modes)
- To be used for 3D cell imaging via tomography



control via frequency tuning

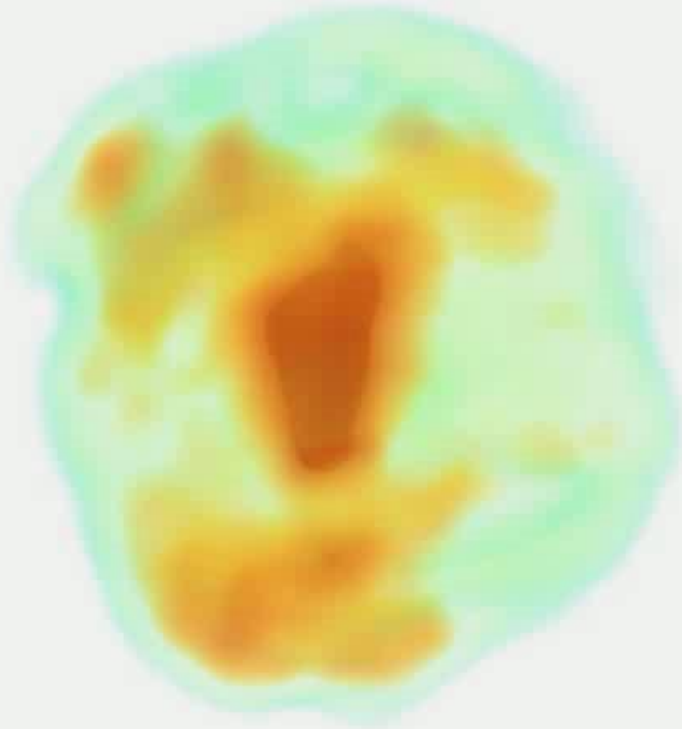


acoustofluidic platform

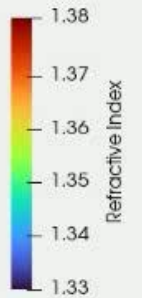
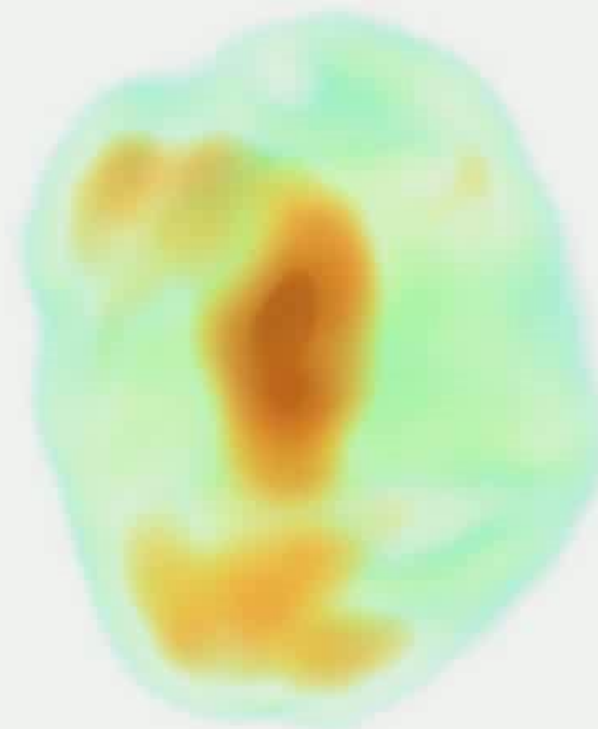


FIRST TOMOGRAPHY RESULT

DHM



Coded WFS



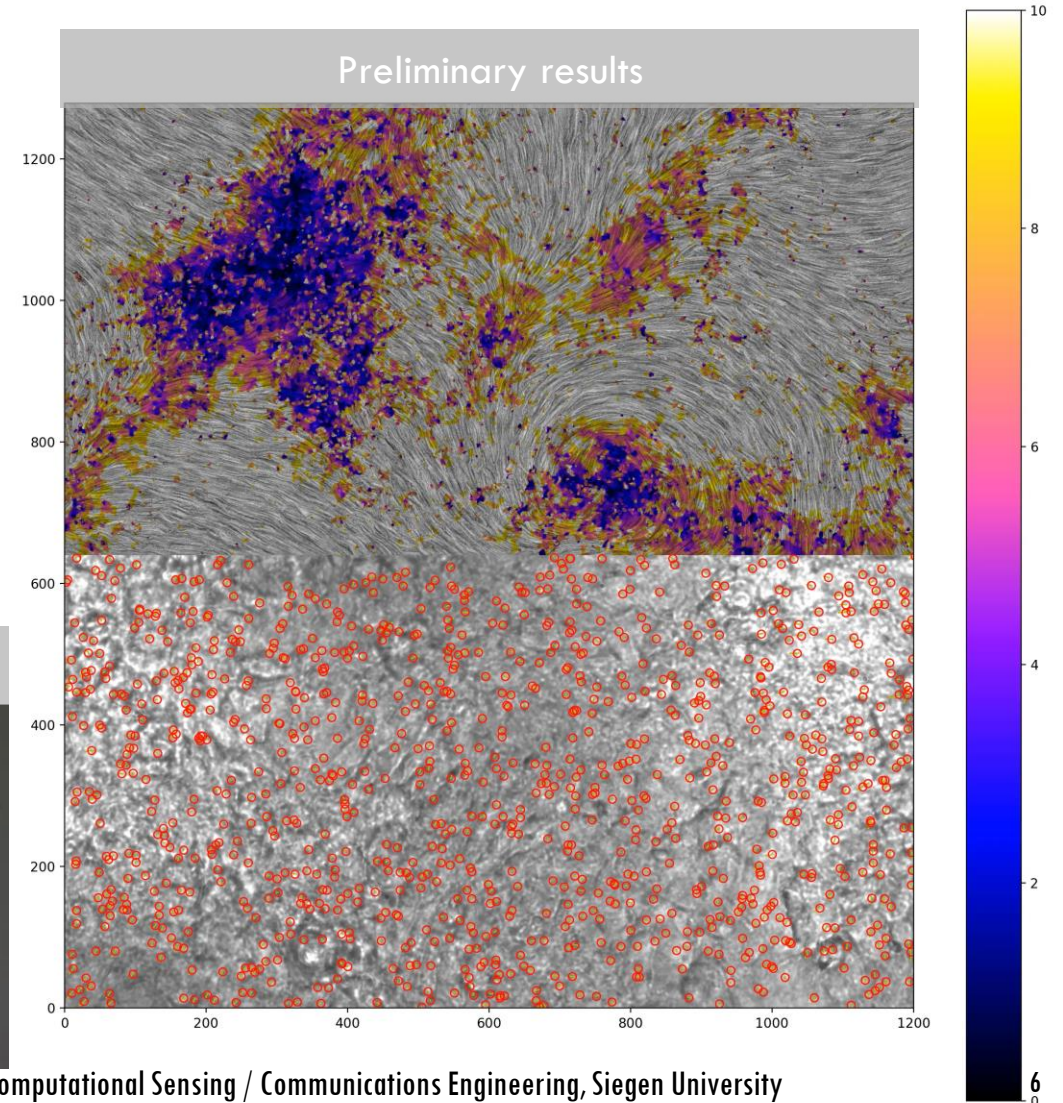
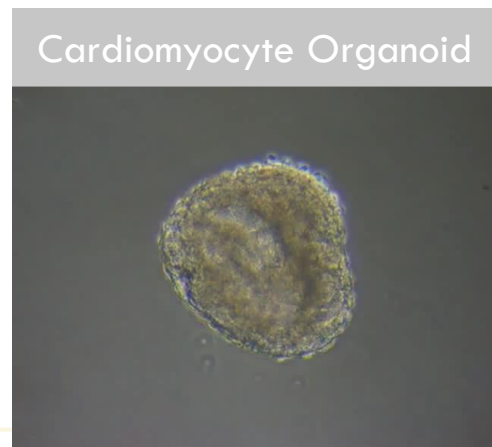
CASE II: ELASTIC DEFORMATION

- Working with a biomedical group in Tampere / Finland

- Target: cardiomyocytes (cardiac muscle cells)
 - These cells show dynamic deformations
 - Different complexity levels:
 - Monolayers
 - Multi-layers
 - Organoids

- Cyclic motion

Show Tampere video



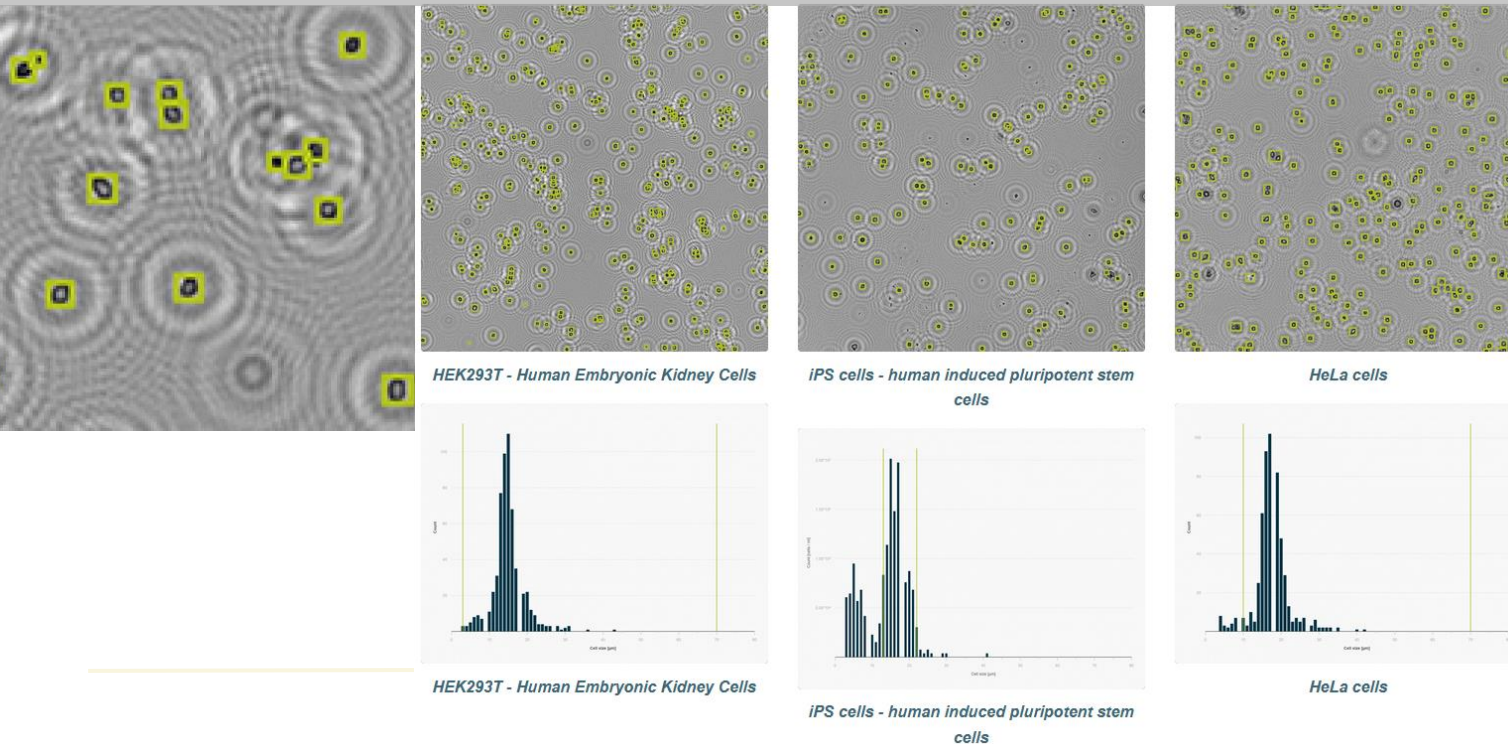
CASE III: HOLOGRAPHIC PARTICLES

- In cooperation with anvaajo GmbH (Dresden)
 - Inline-holographic images of particles
 - Goal: 3D tracking (tumbling particles would yield 3D projections enabling improved classification)
 - Constraints: random walk, image formation: coherent imaging

Point of care testing device for veterinarians



Example cells in urine



- Red Blood Cells/ White Blood Cells
- Squamous Epithelial Cells/ Non-Squamous Epithelial Cells
- Hyaline Casts/ Non-Hyaline Casts
- Calcium Oxalates Dihydrates/ Struvites/ Unclassified Crystals
- Bacteria (suspected presence)

WORK PLAN

Monat	1-3	4-6	7-9	10-12	13-15	16-18	19-21	22-24	25-27	28-30	31-33	34-36
AP 1	AP 1.1			AP 1.2					AP 1.3			
AP 2.1	Fall I				Fall II				Fall III			
AP 2.2	Fall I				Fall II				Fall III			
AP 2.3		Fall I				Fall II				Fall III		
AP 3	AP 3.1				AP 3.2							
AP 4	AP 4.1				AP 4.2				AP 4.3			
AP 5	AP 5.1					AP 5.2						

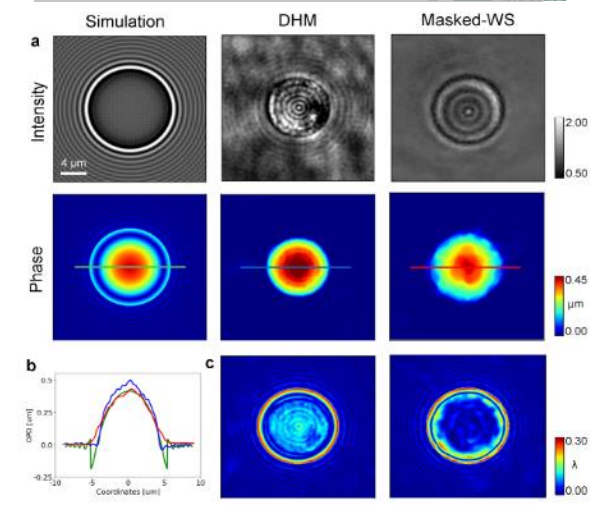


Fig. 1. Accuracy validation measurements using a 10 μm spherical silica bead. (a) The first and second rows show intensities and phases (in OPD) method-wise, respectively. (b) OPD of a cross-section profile of the bead relative to the immersion. (c) The RMSE of the DHM (left) and Masked-WS (right) OPDs from the phase of the simulated bead in (a), normalized by the wavelength λ_{III} .

- **Current state:**

- Data availability: case I and case II, data available
- Ground truth: simulation of transparent objects in microscopes is available
- Classical methods: case I and case II have partial results

AP2	Anwendung Mikroskopie, Koordination: USi2	Insgesamt 41PM
PM/Partner	36PM USi2, 5PM UMa	
AP2.1 Datenaufbereitung jeweils für Fall I,II) und III), AP2.1.(I-III) klassische Methoden und Annotation (6PM USi2)		
AP2.2 Datensimulation für Fall I,II) und III), Wellenoptische Simulationen mit entsprechenden Randbedingungen (6PM USi2)		
AP2.3 Constraintmodellierung für Fall I, II) und III), AP2.3.I: 3D Festkörpertransformation + TIE, AP2.3.II: Loop Closure + Schichtmodell, AP2.3.III Brownsche Bewegung + Holografie (24PM USi2, 5PM UMa)		

Thank you!



CHAIR FOR
COMPUTATIONAL SENSING &
COMMUNICATIONS ENGINEERING
SIEGEN UNIVERSITY

CONTACT: IVO.IHRKE@UNI-SIEGEN.DE